

Vascular Calcification: Where is the Cure?

Wen-Wen Liu, Mei-Lin Liu*

Department of Geriatrics, Peking University First Hospital, Beijing 100034, China

ABSTRACT

With the progress of aging, the incidence of vascular calcification (VC) gradually increases, which is correlated with cardiovascular events and all-cause death, aggravating global clinical burden. Over the past several decades, accumulating approaches targeting the underlying pathogenesis of VC have provided some possibilities for the treatment of VC. Unfortunately, none of the current interventions have achieved clinical effectiveness on reversing or curing VC. The purpose of this review is to make a summary of novel perspectives on the interventions of VC and provide reference for clinical decision-making.

Key words: vascular calcification; clinical; pathophysiology; therapeutic strategies; novel findings

INTRODUCTION

The prominent manifestation of vascular calcification (VC) is the ectopic deposition of calcified plaques caused by the disorder of mineral metabolism, which is associated with senescence and renal dysfunction. Ectopic calcification usually occurs in intima and media of the artery of communications and micrangium as well as in the heart valves and calciphylaxis. VC is common in chronic kidney disease (CKD), especially in patients with end-stage renal disease (ESRD). Coronary arterial calcification (CAC) significantly increases the risk of cardiovascular events, including myocardial infarction, heart failure, and cardiac death in patients with CKD^[1]. Clinical studies have shown that current attempts on reversing VC are insufficient to obtain satisfactory effects. The mechanism of VC is so complex that effective prevention and treatment strategies remain limited.

The review outlines strategies to treat VC and its pathogenesis, emphasizing the importance of managing secondary hyperparathyroidism and cardiovascular risk factors. Inhibition of vascular smooth muscle cell transformation and calcium deposition are crucial drug

targets. Stem cell technology and drugs like SNF472 show potential as therapies, offering new insights into VC research and treatments.

PATHOPHYSIOLOGY OF VASCULAR CALCIFICATION

Previous studies have manifested that VC is caused by hyperphosphatemia and other risk factors including oxidative stress, inflammation, apoptosis, and lipid deposition^[2]. VC refers to the pathological deposition of hydroxyapatite in the vascular wall, which is closely attributed to the imbalance of calcium and phosphorus metabolism. In addition, calcified blood vessels continue to mineralize, similar to the ossification process in bones. Therefore, even the calcium and phosphorus disorders have been restored, the existing calcification will continue to progress, which is the main challenge in calcification treatment. *In vitro* primary vascular smooth muscle cells (VSMCs) are possible to be induced to undergo phenotypic transformation expressing osteogenic markers and also lose their specific markers.

Intimal calcification is mainly caused by metabolic diseases, which is characterized by the deposition of lipids and lipoproteins in the intima of large and medium arteries, leading to luminal stenosis, which is similar to the pathogenesis of atherosclerosis. Medial calcification is the main manifestation of VC in CKD patients, which is characterized by the linear deposition of hydroxyapatite in the arterial media,

Received April 18, 2024; accepted June 28, 2024; published online September 4, 2024.

*Corresponding author E-mail: liumeilin@hotmail.com.

© The authors 2024. Published by Chinese Academy of Medical Sciences. This is an open access article attributed under the CC BY-NC license (<http://creativecommons.org/licenses/by-nc/4.0>).

resulting in decreased vessel wall compliance, increased pulse pressure, and secondary ischemia, heart failure, and stroke. Heart valve calcification is also common in CKD patients. Studies have shown that calcium deposition in the heart valves is increased in dialysis patients, mainly involving the aortic and mitral valves, which significantly increases the risk of cardiovascular events in CKD patients. Calciphylaxis, also known as calcific uremic arteriopathy, is commonly seen in patients with ESRD. It is a lethal condition characterized by ischemic necrosis of tissues and organs due to calcification, intimal hyperplasia, and thrombosis in systemic small arteries^[3]. The disease is mainly characterized by extremely painful skin ulcers, and wound eschar formation can also occur in lung, digestive system, penis, and other organs, manifested as unexplained recurrent dyspnea and gastrointestinal bleeding^[4,5]. At present, no unified diagnostic criteria for calciphylaxis have been available. Due to its insidious onset and lack of clinical understanding, calciphylaxis is often diagnosed at an advanced stage, so the treatment is difficult and the prognosis is extremely poor.

Senescence-related mitochondrial damage or dysfunction leads to insufficient energy required for physiological operation of all cells, which is widely recognized as a profound mechanism of VC. More than two-thirds of patients over 70 years of age have extensive VC, and more than 96% of these elderly individuals exhibit CAC^[6].

THERAPEUTIC AGENTS FOR VASCULAR CALCIFICATION

Vitamins

Vitamin D

Vitamin D is a lipid-soluble vitamin that can promote the absorption of calcium by the small intestinal mucosa to increase the circulating calcium and phosphorus and regulate bone health. Vitamin D also promotes cell growth and differentiation, regulates immune system, and affects inflammatory response. 1,25-hydroxyvitamin D₃ (1,25(OH)₂D₃, or calcitriol), the active form of vitamin D, can induce the synthesis of calcium-binding protein that is required in calcium absorption in the intestine and promote the renewal of calcium and osteogenesis^[7]. Furthermore, calcitriol also promotes the reabsorption of calcium and phosphorus in renal tubules^[7].

The effect of vitamin D supplementation on VC re-

mains controversial. Numerous studies suggest that vitamin D excess increases the risk of VC. Other studies indicate that vitamin D deficiency also leads to VC, and long-term vitamin D supplementation is recommended to prevent calcification. The inconsistent conclusions may stem from two aspects: on the one hand, vitamin D increases circulating concentrations of calcium and phosphate by promoting intestinal and renal absorption. Animal experiments have demonstrated that vitamin D significantly accelerates arterial calcification, but vitamin D doses in these experiments tended to be very high (100,000 to 500,000 IU/kg/day) which is close to toxic dose^[8]. Vitamin D, on the other hand, may have anti-inflammatory effects, which have potential benefits for vascular diseases^[7]. In several large clinical and epidemiological studies, vitamin D deficiency was significantly associated with cardiovascular and non-cardiovascular mortalities^[9]. One clinical study showed that patients with either low or high vitamin D had greater carotid thickness and calcification than patients with normal levels^[10]. Due to the rarity of vitamin D excess in the population and the ethical violation of inducing vitamin D toxicity in patients, the relationship between vitamin D excess and VC cannot be conducted in clinical studies. Based on the current evidence, the effect of vitamin D excess on promoting calcification may be less prominent than that of vitamin D deficiency.

Vitamin K

Vitamin K is a fat-soluble vitamin that has two natural analogues: vitamin K₁ (chloroquinone) and vitamin K₂ (mequinone, MKs). Vitamin K can be obtained from diets, synthesized by intestinal bacteria and artificial supplements, and transported through the lymphatic system. Studies indicate that, compared with vitamin K₁, vitamin K₂ has a more comprehensive effect on human body^[11]. A total of 17 vitamin K-dependent proteins (VKDPs) have been discovered, which play important roles in regulating mineral metabolism, osteogenesis, VC, and inflammatory responses. The primary function of vitamin K is to serve as a coenzyme of coagulation factor (gamma-carboxylase) and an essential substance for the synthesis of clotting factors II, VII, IX, and X^[12]. Vitamin K deficiency may prolong blood clotting time and generate severe bleeding.

Matrix Gla protein (MGP) is mainly synthesized in chondrocytes and VSMCs, which are the primary suppressant of calcinosis, assisting to prevent VC^[13]. After translation in the cytoplasmic endoplasmic reticulum,

MGP is rapidly carboxylated by gamma-glutamyl carboxylase (GGCX). This process requires vitamin K as a key cofactor to form the carboxylated MGP. Secreted by osteoblasts, osteocalcin (alternative name is bone gla protein) is a component that promotes calcium deposition into bone^[14]. Growth arrest-specific protein 6 (Gas6), a component of the tyrosine kinase receptor family, inhibits VC by suppressing apoptosis of endothelial cells and VSMCs^[15]. Gla-rich protein has been regarded as a new type of VKDP in sturgeon cartilage, which contains many Gla residues and may be a potential inhibitor of VC^[16]. Therefore, vitamin K can inhibit the progression of VC by facilitating the activities of VKDPs.

Vitamin K alleviates lipopolysaccharide-induced inflammatory responses by intercepting the transduction of nuclear factor κ B (NF- κ B) signaling pathway^[17]. Vitamin K level is suggested to have a negative association with certain inflammatory markers such as interleukin-6 and C-reactive protein^[18]. However, a randomized trial recruiting 379 healthy subjects receiving vitamin K₁ supplements for three years failed to observe any changes in inflammatory biomarkers^[19]. A multicenter trial enrolled 365 elderly men with an aortic valve calcification (AVC) score >300 arbitrary units (AU) manifested that two-year MK-7 plus vitamin D supplementation made no difference on AVC progression^[20]. A phase II trial to evaluate the influence of vitamin K supplementation over one-year follow-up on vascular stiffness or calcification in kidney transplant recipients also received a disappointing result^[21].

Statins

Statins are the acknowledged basic drugs for the prevention and treatment of coronary heart disease by competitively inhibiting the rate-limiting enzyme hydroxy methylglutaryl coenzyme A (HMG-CoA) of endogenous cholesterol synthesis, blocking intracellular cholesterol synthesis, and effectively reducing total cholesterol and low-density lipoprotein. In recent years, studies reveal that statins have a variety of other non-lipid-lowering properties, including stabilizing atherosclerotic plaques, treating osteoporosis, anti-inflammation, anti-oxidation, anti-tumor, and anti-Alzheimer's disease. A series of imaging studies have shown that high-intensity statin therapy is associated with plaque volume reduction by reducing fatty and necrotic plaque ingredients and increasing plaque calcification. Statins have anti-inflammatory and antioxidant properties in addition to their cholesterol-lowering effects. Ivanovski *et al.*^[22] first demonstrated

that simvastatin could reduce oxidative stress (OS) markers and aortic calcification in apolipoprotein E deficient mice with CKD, independent of cholesterol changes. Further studies indicated that pravastatin was capable of decreasing OS level through inhibiting farnesyl transferase in calcified VSMCs^[23]. Statins could increase atherosclerotic calcification by disinhibiting the Rac1-IL-1 β signaling axis of macrophage^[24]. Statins and vitamin K₂ can inhibit apoptosis-mediated VSMC calcification by activating Gas6/Axl-PI3K/Akt pathway^[25,26]. After five years of follow-up, a population-based study implied that statin treatment was independently correlated with CAC progression^[27]. An analysis of two randomized controlled trials comparing the influence of high-dose statin therapy (atorvastatin 80 mg) and low-to-moderate-intensity statin therapy (pravastatin 40 mg or atorvastatin 10 mg) on CAC failed to demonstrate any difference in Agatston score or CAC volume within one year^[28]. However, the available data have not reached a uniform conclusion about the relationship between statin use and VC progression.

Matrix Gla protein

Matrix Gla protein (MGP) is a vitamin K-dependent calcium-binding protein synthesized by osteogenesis and VSMCs. It is a carboxyglutamic acid translated in bones, cartilage, heart, and kidneys. This protein acts as an inhibitor of vascular mineralization and plays a role in bone tissue. Keutel syndrome is a rare autosomal recessive disorder characterized by severe calcification of soft tissues, which is thought to be caused by mutations in the gene encoding for MGP^[29]. The possible mechanisms that MGP inhibits calcification can be summarized in two aspects. First, MGP has a high affinity for calcium ions and hydroxyapatite crystals, which can be used as a chelating agent^[30]. Second, MGP reduces the ability of osteoblast differentiation by binding bone morphogenetic protein-2 (BMP-2)^[31]. The ability of MGP inhibiting calcification is not limited in bone metabolism but also in atherosclerotic calcified plaques and calcified lesions of vascular media^[32,33]. Therefore, MGP is considered to be a crucial suppressant of the intima and media of VC. However, there is no clinical evidence of MGP for the treatment of VC.

Mitochondrial function modulators

Nicorandil is the first clinically used mitochondrial ATP-sensitive K⁺ channel activator with abilities to prevent intracellular Ca²⁺ release and stimulate K⁺ internal

flow, thus maintaining membrane integrity and energy metabolism^[31]. As impaired mitochondrial function is partially related to VC in patients with CKD, nicorandil can relieve VC in rat models of CKD. Nicorandil reversed mitochondrial function in adenine-induced CKD rats by enhancing mitochondrial depolarization and energy metabolism, and reducing mitochondrial swelling^[34,35]. In addition, nicorandil has been proved to facilitate antioxidant capacity and inhibit reactive oxygen species (ROS) production^[35], resulting in reduced aortic calcification^[34,35]. Clinical studies have confirmed that nicorandil is suitable for all types of angina pectoris and can significantly reduce the risk of cardiovascular events. However, the protective effect of nicorandil against myocardial reperfusion injury was not present in potentially calcified rat hearts^[36].

Mitochondrial autophagy is an evolutionarily conserved biological process of mitochondrial degradation and recycling. Under the stimulation of ischemia and hypoxia in mitochondria, the activity of electron transport chain is weakened, resulting in the obstruction of ATP production, the disappearance of mitochondrial membrane ion gradient and Ca²⁺ overload, and finally the opening of mitochondrial membrane permeability conversion pores and the entry of cytochrome c and various mitochondrial proteins into the cytoplasm, thus inducing cell apoptosis. Activation of mitochondrial autophagy can prevent the accumulation of damaged mitochondria and protect VSMCs from oxidative stress, inflammatory response and apoptotic cascade reaction. However, excessive mitochondrial autophagy can lead to repeated degradation of normal mitochondria, affect energy production, lead to cell death, and eventually cause serious damage to tissues and organs. Autophagy activation in the presence of cell damage is thought to be protective, while autophagy defects in normal cells lead to apoptotic activation. Therefore, it is of great significance to regulate proper mitochondrial autophagy to ensure its cell-protective function. As a key regulator of mitochondrial division, mitochondrial dynamin-related protein 1 (Drp1) plays an important role in triggering mitochondrial autophagy. Failure of mitochondrial autophagy can activate Drp1-mediated apoptosis pathway, promote vitamin K₂-induced osteoblast differentiation and bone mineralization, and eventually affect VC^[37]. The interaction between mitochondrial autophagy and VC is complex and can be influenced by disease progression, anatomical location, and surrounding microenvironment. While

the clinical role of Drp1 in VC is still unclear, inhibiting Drp1 is a potential target to treat VC.

Hydroxyapatite inhibitors

The phosphate binder sevelamer^[38] and the cinacalcet^[39], for the treatment of hyperphosphoremia and secondary hyperparathyroidism, reduce the potential risk factors for VC in ESRD, but they can hardly resolve the biological issues responsible for this complex pathological phenomenon. Myo-inositol hexaphosphate (IP6, phytate), a substance from fiber-rich foods such as grains, nuts, and legumes, has been applied to treat cardiovascular calcification^[40,41]. The mechanism of IP6 is completely different from that of the above-mentioned drugs. IP6 binds to hydroxyapatite in a concentration-dependent manner, preventing the formation and aggregation of hydroxyapatite crystals and fundamentally inhibiting the formation of calcification products. Although IP6 has a natural vascular protective effect, its clinical application is limited by poor oral bioavailability^[42].

SNF472 was developed as an intravenous formulation of IP6 to intervene calciphylaxis and VC in hemodialysis patients, and it can be administered via the hemodialysis line to achieve direct inhibition of hydroxyapatite aggregation without being easily cleared by dialysis^[43,44]. SNF472 can inhibit the occurrence and development of ectopic calcification, thereby preventing the formation of hydroxyapatite in blood vessels and treating VC. A phase I clinical trial proved that in hemodialysis patients, treatment with SNF472 9 mg/kg inhibited the hydroxyapatite by 80% (compared with 9% in the placebo group) without affecting the serum calcium level and did not cause clinical symptoms related to hypocalcemia^[45]. Valve replacement is the only approach that has been proven to improve membrane function. There is no effective drug that can delay the progression of valvular disease. CaLIPSO (Ca for calcium and IP6 meaning the item itself)^[46] is the first multicenter clinical trial to compare progression of CAC between 52-week administration of SNF472 and placebo. Results showed that SNF472 significantly alleviated CAC and aortic valve calcification in hemodialysis patients apart from fundamental treatments. The CaLIPSO further evaluated the changes in bone mineral density (BMD) of the total hip and femoral neck and found that BMD decreased in all groups during the follow-up period, but there was no correlation between changes in the vol-

ume of CAC and changes in BMD at other locations, and fracture events were rare^[47]. SNF472 also had no significant effect on serum alkaline phosphatase, calcium, magnesium, phosphate, and intact parathyroid hormone, which are related to bone metabolism. Preliminary findings from studies of SNF472 in animal models of cardiovascular calcification and in patients undergoing dialysis have led to the advancement of this compound into phase II clinical trials for the treatment of calciphylaxis^[48]. However, the study was a single-arm, open-label trial with a small sample size that was not sufficient to provide a clinically significant result. SNF472 has entered phase III clinical trial, and preliminary results suggested that it might become the first drug approved to cure VC or calciphylaxis^[49].

Antioxidant therapies

Oxidative stress (OS) is a vital inducement of cellular aging, which may cause severe damage to cellular organelles and nucleic acids, proteins, and cytomembranes. ROS production in calcified VSMCs may cause telomere shortening, mitochondrial DNA damage, and gene mutations, resulting in VSMC senescence and VSMC transformation. These sequence of changes will activate osteoblast transcription^[50].

Coenzyme Q10 is one of the substances involved in the electron transport chain and aerobic respiration in eukaryotic cell mitochondria. It can promote oxidative phosphorylation and protect the integrity of cytomembranes. Coenzyme Q10 is widely used in the treatment of viral myocarditis, chronic heart failure, hepatitis, and cancer, with an ability to reduce the adverse reactions caused by radiotherapy and chemotherapy^[51]. In animal models with CKD, coenzyme Q10 was found to suppress ROS production and reduce apoptosis, thus alleviating VC^[52]. Moreover, administration of coenzyme Q10 could reduce aortic stiffness in the elderly by alleviating mitochondrial ROS, compared with placebo^[53].

Alpha-lipoic acid (ALA) is a natural antioxidant that has multifarious functions. Additionally, ALA is suggested to enhance mitochondrial function and inhibit cell apoptosis^[54]. ALA can be used in the clinical treatment of acute and chronic hepatitis, liver cirrhosis, hepatic coma, fatty liver, diabetes, skin aging, and other diseases. In *in vivo* models of CKD, ALA attenuated VC by inhibiting ROS and enhancing mitochondrial function through regulating the Gas6/Axl/Akt pathway to reduce apoptosis^[55]. Nonetheless, treat-

ment with ALA 600 mg/d for two to six months failed to change the OS or inflammatory markers in both nondialysis and hemodialysis CKD patients, compared with placebo^[56,57].

Sodium thiosulfate (STS) has antioxidant capacity and direct chelation of calcium that commonly used to treat VC^[58]. STS is found to inhibit BMP-2 synthesis and promote MGP function in calcified VSMCs^[59], suggesting that STS might be involved in the formation of VC. In adenine-induced CKD rats, STS reduced VC without improving mitochondrial function or OS^[35]. STS combined with other antioxidants or mitochondrial function modulator may provide a synergistic effect in VC intervention. STS combined with nicorandil effectively restored mitochondrial metabolism, improved antioxidant capability, reduced ROS production, and ultimately alleviated VC^[35]. With the help of fat cells, STS could alleviate arterial calcification to a greater extent^[60]. These results suggest that the role of STS in reducing VC may be attributed to direct chelation of calcium instead of antioxidant capacity. Moreover, STS treatment has been proved to reduce arterial stiffness, stabilize VC, and prevent heart valve calcification in hemodialysis patients^[61,62].

Quercetin, a natural and dietary source of flavonol, has antioxidant and antiinflammatory activities by inhibiting mitogen-activated protein kinase (MAPK) and NF- κ B pathways^[63]. An optimized dose of quercetin can reduce VC through inhibiting ROS, enhancing ATP synthesis, and alleviating apoptosis in animal models with CKD^[64]. Quercetin mitigates aortic calcification and chondrogenesis through suppressing the expression of transglutaminase 2 and β -catenin^[65]. Additionally, quercetin can partially decrease aortic calcification through enhancing the level of superoxide dismutase 2 and activating nitric oxide synthetase (iNOS)/MAPK pathway^[66]. Nevertheless, the phase II clinical trial manifested that quercetin combined with sodium nitrite failed to optimize vascular function in predialysis CKD patients compared with placebo, including biomarkers of endothelial function (*e.g.*, vascular adhesion molecule, e-selectin, and von Willefamer factor), inflammation, and OS^[67].

Anti-inflammatory therapies

Systemic and local inflammation play a key role in the occurrence and development of VC. In addition to mineral metabolism imbalance, the oxidative stress, inflammation, and cell signal transduction

abnormal factors are also involved in VC. Anti-inflammatory therapies contribute to reduce the production of inflammatory cytokines (*e.g.*, IL-6 and tumor necrosis factor alpha [TNF- α]) and C-reactive protein during inflammation, which promotes the transformation of VSMCs into osteoblast-like cells. The use of anti-inflammatory drugs, including non-steroidal anti-inflammatory drugs (NSAIDs), glucocorticoids, or more targeted biologic agents, can reduce the production of these inflammatory mediators and slow the calcification process. The accumulation and activation of inflammatory cells such as macrophages and lymphocytes at the site of VC can exacerbate local inflammatory responses. Anti-inflammatory therapies can inhibit the activation and migration of inflammatory cells and reduce inflammation of vascular injuries. Receptor activator of nuclear factor-kappa B ligand (RANKL)/receptor activator of nuclear factor-kappa B (RANK)/osteoprotegerin (OPG) passway associated with bone remodeling also plays a key role in VC. Certain anti-inflammatory drugs may slow down the process of calcification by inhibiting VSMC ossification through regulating bone formation correlation factors. Denosumab, a specifically targeted monoclonal antibody against RANKL, reduces disease progression in patients with aggressive fibrous dysplasia of the bone and treats osteoporosis. Clinical studies have shown that denosumab has the potential in reducing VC^[68], but not in another trial^[69]. Given the small sample sizes, no definite conclusions have been available.

Monoclonal antibodies, particularly those targeting specific inflammatory factors, provide a new insight for VC treatment. For instance, the TNF- α inhibitors (infliximab or adalimumab) reduce inflammation through direct combination with excessive TNF- α , which may be beneficial to controlling VC. Animal studies have shown that the infliximab could block the aorta BMP-2, Msx2, Wnt3a, and Wnt7a signaling pathways and significantly reduce the aortic calcium accumulation^[70]. However, there is almost no relevant clinical research^[71]. IL-6 is an essential mediator of inflammation and autoimmune diseases. The inhibitor (tocilizumab) can effectively reduce inflammation, which may be conducive to inhibiting VC. With a deeper understanding of the inflammatory mechanisms of VC, more biological agents targeting specific inflammatory pathways such as IL-1 β and IL-17 are being developed, aiming to intervene in the

inflammatory process more precisely. Although the anti-inflammatory and immune therapies have shown certain potentials, their actual efficacy, long-term safety, and optimal timing of use in VC treatment warrant more studies.

Mineral metabolism regulators

The balance of calcium and phosphorus metabolism is extremely essential for the organs and cells to maintain normal physiological functions. CKD patients often also have metabolic disorders of minerals such as calcium, phosphorus, and magnesium. Severe disturbances in calcium and phosphorus homeostasis, particularly hyperphosphatemia, are recognized as pivotal contributors to the pathogenesis of arterial calcification and the onset of other cardiovascular complications in CKD individuals. Phosphate is related to vascular inflammation and can induce VSMCs to produce a variety of proinflammatory cytokines such as tumor necrosis factor, IL-1 β , and IL-6. Oxidative stress induced by inflammation is a strong inducer of VSMC osteogenic transformation and VC. The increase in calcium and phosphorus shows a synergistic effect in inducing VC. Calcium-induced VC may be achieved by promoting the release of VSMCs matrix vesicles and cell apoptosis. In addition, hypercalcemia is an independent risk factor for aortic calcification in CKD patients. Secondary hyperparathyroidism is also involved in the occurrence of VC. A rat model of uremia demonstrated the relative effect of high blood phosphorus and high parathyroid hormone on VC. It was found that high parathyroid hormone had a direct effect on VC and could also enhance the VC induced by high phosphorus. Stabilization of calcium and phosphorus levels in patients with CKD is particularly critical for preventing VC progression and prolonging life. Servilam, a calcium-free phosphate binder, reduces treatment-associated hypercalcemia, but its effects on reducing VC and all-cause mortality have not been conclusively concluded^[72,73]. Calcimimetics is used as a quasi-calcium agent that can reduce serum parathyroid hormone, phosphorus, and calcium levels in patients with secondary hyperparathyroidism^[74,75].

In addition to traditional calcium and phosphorus, magnesium may be associated with the incidence of cardiovascular diseases in patients with CKD. Magnesium, a cofactor of various metabolic reactions in human body, is involved in regulating ion channel cur-

rents. It plays a momentous role in regulating cell growth and maintaining the excitability of myocardium and skeletal muscle. Recent evidence suggests magnesium is involved in many cardiovascular function-regulating processes, and disruption of magnesium homeostasis can precipitate a spectrum of cardiovascular pathologies including hypertension, arrhythmia, atherosclerosis, arterial calcification, and vascular aging. Magnesium ions can activate the expression of calcium-sensitive receptors and are considered to be natural physiological calcium blockers. Magnesium deficiency promotes VC. Combined with the use of quasi-calcium, magnesium supplementation can reduce the deposition of minerals in blood vessels^[76]. In animal studies, magnesium supplementation has been observed to repress the expression of osteogenic genes, thus suppressing the occurrence of calcification^[77]. It has also been proved that magnesium can regulate calcification by affecting calcium and phosphorus metabolism, lipid deposition, oxidative stress, inflammation, and other processes. Magnesium supplementation may be a new therapeutic approach for delaying or alleviating VC. However, multiple randomized controlled trials have shown that magnesium supplementation alone is inadequate to delay the progression of VC, and other treatment strategies are badly in need to reduce the risk of cardiovascular disease in patients with CKD^[78,79].

POTENTIAL TARGETS IN VASCULAR CALCIFICATION

Iron homeostasis

Some new molecular targets provide new perspectives and possibilities for the treatment of VC. Iron is a pro-oxidant that can induce ROS productions, and its overload can aggravate oxidative stress and VC^[80]. Ferroptosis is a type of cell death characterized by the accumulation of supersize oxidative stress products. Iron overload is associated with a variety of diseases, including cardiovascular diseases. Ferroptosis occurs in VSMCs during calcium-phosphate-induced VC, and inhibiting iron death significantly reduces osteogenic differentiation and calcification of VSMCs. Regulating iron homeostasis, such as lowering excess iron in the body through chelating agents, may help slow or prevent the process of VC. Deferoxamine chelates free or protein-bound 3-valent iron (Fe^{3+}) to form a stable, non-toxic, water-soluble iron-amine complex that is

excreted via urine. Primarily, it is used for relieving acute iron poisoning. Animal studies have shown that deferoxamine can reduce aortic calcification in rats with CKD. Therapeutic strategies targeting specific iron metabolism in VC are still under investigations^[81].

Epigenetic modification

The transdifferentiation of VSMCs from contractile type to osteoblasts is the key molecular event of VC. Epigenetic modification is involved in the regulation of target gene expression through DNA methylation, histone modification, and non-coding RNA regulation without changing the DNA coding sequence. It can regulate the transdifferentiation of VSMCs and participate in the regulation of VC. Specific epigenetic modification enzymes such as demethylase alkB homolog 1 (ALKBH1) are involved in the regulation of VC^[82]. Upregulation of *ALKBH1* gene leads to decreased demethylation level, and *ALKBH1* overexpression aggravates the progression of VC. Targeting *ALKBH1* may be a therapeutic approach to reducing VC burden in CKD patients^[83]. Sirtuin 6 (SIRT6), a member of the Sirtuin family, is a class III histone deacetylase and a key epigenetic regulator. SIRT6 inhibits osteogenic transdifferentiation of VSMCs by down-regulating human runt related transcription factor 2 (RUNX2) and plays a role in alleviating VC. Thus, SIRT6 may be an attractive therapeutic target for VC^[84]. Non-coding RNAs, such as miRNAs, affect the osteogenic differentiation of VSMCs and regulate VC by affecting the expression of osteogenic factors such as RUNX2 and alkaline phosphatase. However, relevant clinical evidence is lacking. Enhancing or inhibiting specific epigenetic modifications to restore normal function of VSMCs may provide new pathways to reduce calcification. More research in these emerging fields is in need of transitioning from basic research to clinical applications.

Stem cell technology

The role of stem cell technology in VC is a cutting-edge and highly concerned research field. Stem cells, especially mesenchymal stem cells (MSCs), have multidirectional differentiation potentials and strong paracrine effect. They can secrete a variety of growth factors and anti-inflammatory factors, which help to promote vascular repair and reduce macrophage-mediated damage^[85]. Stem cells may be involved in remodeling damaged vascular tissue by inducing differentiation into vascular smooth muscle cells or endothelial cells^[86]. Au-

tologous or allogeneic stem cells can be injected into diseased vascular region with the expectation that they would differentiate into functional vascular cells to alleviate or reverse calcification. In addition to direct transplantation, stem cells might also be used as a vector carrying anti-VC gene to achieve more precise therapeutic effect.

Ectoblastic cells expressing specific adhesion molecules of stem cells, including Sca1 and/or CD34, has the potential to differentiate into VSMCs^[87]. Mesenchymal stem cell (MSC)-like cells, specifically those positive for Gli1 expression, express a number of known vascular progenitor cell markers, including CD34, Sca1, and platelet-derived growth factor receptor beta (PDGFR β). Gli1+ cells are located in the outer membrane of the artery and are progenitors of VSMCs, contributing to intimal recovery and regeneration after acute injury. Gli1+ cells are the main source of osteoblasts during VC. Gene modification of Gli1+ cells prior to renal injury significantly reduced the degree of VC^[88], suggesting that Gli1+ cells are vital adventitial progenitor cells in vascular remodeling after acquired injuries, and they might be the most promising therapeutic targets for alleviating VC. However, these theories are based on animal experiments, and there is no evidence for clinical application.

The current drugs related to the treatment of VC and the relevant clinical and preclinical studies are listed in **Table 1**.

INTERVENTIONAL THERAPIES IN VASCULAR CALCIFICATION

CAC lesions bring great difficulties to interventional therapies, which increases the risk of surgical failures (e. g., inadequate balloon or stent dilation), complications (e. g., coronary dissection, coronary perforation, or balloon rupture), and perioperative mortality^[89]. A variety of techniques have been developed to address the diverse challenges posed by calcified lesions, including various balloons, rotational, orbital and laser atherectomy, and intravascular lithotripsy (IVL)^[90].

Balloon angioplasty (BA) is an interventional cardiovascular treatment technique that is mainly used to treat stenosis or occlusion of coronary artery, carotid artery, renal artery, lower extremity artery, and other vessels. The procedure involves making a small incision in the skin, delivering a catheter with an inflat-

able balloon to the site of the diseased blood vessel, and then using pressure to expand the balloon to squeeze and flinch the atherosclerotic plaque on the inner wall of the blood vessel, thereby restoring blood flow. Non-Compliant (NC) balloons are often applied in the treatment of moderate CAC to achieve more ideal stent dilation that can withstand high pressure and achieve more uniform balloon dilation. BA with NC balloon is an auxiliary therapy performed after atherectomy to ensure sufficient plaque modification prior to stent releases^[89]. High-pressure balloon is a NC balloon catheter that can withstand high pressures (up to 35 atm) without significantly changing the diameter inside the balloon^[91]. Cutting balloon is another NC balloon catheter with the blades making shallow incisions in calcified plaques to facilitate stent implantation. The microsurgical blades are mainly applied in ostial or in-stent restenosis (ISR) lesions^[92]. Scoring balloon is surrounded by sharp scratch elements, allowing concentrated force into calcified plaques throughout the expansion process. Compared with cutting balloons, scoring balloons reduced the risk of vascular wall injury and coronary dissection^[93].

Intravascular lithotripsy (IVL) is a novel technique for treating severe CAC. The shock waves produce positive pressures (up to 50 atm) to selectively rupture the surface and deep layers of the calcified regions without affecting the surrounding soft tissues^[94]. IVL is administered safely without causing coronary dissection, slow flow phenomenon, embolism, or coronary perforation^[95-98].

Rotational atherectomy (RA) mechanically ablates hard calcified stenosis with a rotating diamond tip while deflecting flexible non-calcified tissues. High-speed burr rotation enlarges lumen, smooths plaque surface, and increases balloon predilation^[89]. Current guidelines indicate that RA improves surgical success in fibrosis or severe CAC (class 2a, evidence B)^[99].

Orbital atherectomy (OA) utilizes a diamond-coated crown to generate targeted force for the ablation of non-flexible calcified plaques. The OA crown is suggested to be performed with a slow and continuous motion in areas that need repetitive ablation. OA generates less slow flow and minor thermal damage during crown orbiting, which is possible to decrease slow flow events and thermal injuries during interventional procedures^[100].

Laser atherectomy modifies plaques by photoablation. This technique produces short wavelengths with high-energy ultraviolet light that causes water

Table 1. Summary of studies on current drug interventions of vascular calcification

Intervention	Study model	Inclusion	Number	Follow-up	Conclusion
Vitamin D	Animal, human	Men from the community with AVC ^[20]	365 ^[20]	24 months ^[20]	Controversial ^[9] , negative ^[20]
Vitamin K	Human	Men from the community with AVC ^[20] , kidney transplant recipients ^[21]	365 ^[20] , 90 ^[21]	24 months ^[20,21]	Negative ^[20, 21]
Statin	Human	Subjects from the HNR study without CAD at baseline ^[27] , hypercholesterolemic women and men ^[28]	3,483 ^[27] , 852 ^[28]	5 years ^[27] , 24 months ^[28]	Negative ^[27,28]
Matrix Gla protein	Molecular, cellular	-	-	-	Positive ^[29,31]
Nicorandil	Animal	-	-	-	Controversial ^[34,36]
SNF472	Animal, human	Adult patients with ESRD receiving hemodialysis ^[46]	274 ^[46]	52 weeks ^[46]	Positive ^[44,46]
Coenzyme Q10	Animal, human	Healthy older adults (60-79 years) with impaired endothelial function ^[53]	20 ^[53]	6 weeks ^[53]	Positive ^[52,53]
Alpha-lipoic acid	Animal	-	-	-	Positive ^[55]
Sodium thio-sulfate	Human	Haemodialysis patients with abdominal aorta calcification ^[61]	60 ^[61]	6 months ^[61]	Controversial ^[61]
Quercetin	Animal, human	Patients with predialysis CKD ^[67]	70 ^[67]	12 weeks ^[67]	Positive ^[64] , negative ^[67]
Denosumab	Human	Hemodialysis patients ^[68,69]	58 ^[68] , 30 ^[69]	2 years ^[68] , over 2 years ^[69]	Positive ^[68] , negative ^[69]
Infliximab	Animal	-	-	-	Positive ^[70]
Servilam	Human	Patients with CKD ^[72] , adults with CKD not on dialysis or post-transplant ^[73]	2,802 ^[72] , 2,498 ^[73]	-(34 RCTs) ^[72] , median 9 months (20 RCTs) ^[73]	Positive ^[72] , negative ^[73]
Calcimimetics	Cell	-	-	-	Positive ^[75]
Magnesium	Animal	-	-	-	Positive ^[77]
Deferoxamine	Animal	-	-	-	Positive ^[81]

AVC: aortic valve calcification; HNR: Heinz Nixdorf Recall; CAD: coronary artery disease; ESRD: end-stage renal disease; CKD: chronic kidney disease; RCTs: randomized controlled trials

evaporation, carbon bond dissociation, and molecular vibrations, contributing to plaque elimination and lumen expansion^[101]. It can be used to treat balloon lesions that cannot span or dilate, chronic complete occlusions, debulking vein graft disease, and calcific non-dilatable in-stent restenosis (ISR)^[102,103].

SURGICAL TREATMENTS IN VASCULAR CALCIFICATION

Surgical bypass or endarterectomy represents the rescue treatment for severe vascular stenosis caused by

heavy VC. Endarterectomy is a surgical procedure that is mainly used to treat the disease of intimal lipid plaque deposition, luminal stenosis, or even occlusion of the large and middle arteries due to atherosclerosis. This procedure mainly targets severe stenoses in the carotid, iliac, and femoral arteries. In endarterectomy, the surgeon will cut open the outer tissue of the diseased vessel, directly expose and remove the hardened plaque and thickened intima on the inner wall of the vessel, so as to restore blood flow and reduce the risk of cardiovascular and cerebrovascular events caused by poor blood flow.

A study comparing percutaneous coronary intervention (PCI) and coronary artery bypass grafting (CABG) in patients with severe CAC within ten-year follow-up showed that the calcification was an independent predictor of mortality; however, there was no difference in prognosis between PCI and CABG treatment^[104]. Clinical evidence demonstrated that anatomic factors of VC would affect the risk of carotid angioplasty and were associated with 30-day risk of stroke or death in patients with symptomatic severe carotid stenosis undergoing endarterectomy or angioplasty^[105].

As an invasive treatment, surgery can be associated with perioperative and long-term complications and therefore is not the preferred choice for calcification intervention.

CONCLUSIONS

Based on the possible mechanisms of VC, the therapeutic strategies focalize in the regulation of bone and mineral metabolism disorders. The management of secondary hyperparathyroidism is essential for rectifying the dysregulation of bone and mineral metabolism, which encompasses conditions such as hyperphosphatemia, hypocalcemia, hypercalcemia, and disturbances in parathyroid hormone levels. Oral administration of phosphate binders, calcitriol, antioxidants, and Ca²⁺ mimics can be helpful. Given the association between intimal VC and atherosclerosis, the management of pertinent risk factors — such as dyslipidemia, hypertension, diabetes mellitus, tobacco use, obesity, and physical inactivity — constitutes a pivotal therapeutic strategy for the prevention of cardiovascular incidents.

Currently, no efficient therapy is available to prevent or reverse vascular or valvular calcification. Inhibition of VSMC phenotypic transformation and calcium deposition are the two main strategies for reversing VC. Stem cell therapy is the most promising intervention, and SNF472 has been applied as an orphan drug to treat peripheral artery disease in ESRD.

The advent of multi-omics technologies, epigenetics, single-cell sequencing, and bioinformatics has furnished novel methodologies for elucidating the underlying mechanisms, pinpointing potential therapeutic targets, and repurposing existing medications for the treatment of vascular and valve calcification.

ARTICLE INFORMATION

Competing interests

The authors declare no conflict of interest.

Authors' contributions

Liu WW and Liu ML wrote, reviewed, and edited the manuscript before submission. Both authors confirm the final version of the manuscript.

Funding

This study was supported by the Peking University Baidu Fund (2019BD019).

REFERENCES

- Chen J, Budoff MJ, Reilly MP, et al. Coronary artery calcification and risk of cardiovascular disease and death among patients with chronic kidney disease. *JAMA Cardiol* 2017; 2(6): 635-43. doi: 10.1001/jamacardio.2017.0363.
- Rocha-Singh KJ, Zeller T, Jaff MR. Peripheral arterial calcification: prevalence, mechanism, detection, and clinical implications. *Catheter Cardiovasc Interv* 2014; 83(6): E212-20. doi: 10.1002/ccd.25387.
- Nigwekar SU, Thadhani R, Brandenburg VM. Calciphylaxis. *N Engl J Med* 2018; 378(18):1704-14. doi: 10.1056/NEJMra1505292.
- Shen G, Huang R, Liu B, Kuang A. Sodium thiosulfate in the treatment of lung and breast calciphylaxis: CT and bone scintigraphy findings. *Clin Nucl Med* 2017; 42(11):893-5. doi: 10.1097/RLU.0000000000001837.
- Kang SJW, Madhan K. Gastrointestinal manifestations in a patient with calciphylaxis: A case report. *Case Rep Nephrol Dial* 2019; 9(2):119-25. doi: 10.1159/000502436.
- Lahtinen AM, Havulinna AS, Jula A, et al. Prevalence and clinical correlates of familial hypercholesterolemia founder mutations in the general population. *Atherosclerosis* 2015; 238(1):64-9. doi: 10.1016/j.atherosclerosis.2014.11.015.
- Wang J, Zhou JJ, Robertson GR, et al. Vitamin D in vascular calcification: a double-edged sword? *Nutrients* 2018; 10(5). doi: 10.3390/nu10050652.
- Shobeiri N, Adams MA, Holden RM. Vascular calcification in animal models of CKD: a review. *Am J Nephrol* 2010; 31(6):471-81. doi: 10.1159/000299794.
- Norman PE, Powell JT. Vitamin D and cardiovascular disease. *Circ Res* 2014; 114(2):379-93. doi: 10.1161/CIRCRESAHA.113.301241.
- Shroff R, Egerton M, Bridel M, et al. A bimodal association of vitamin D levels and vascular disease in children on dialysis. *J Am Soc Nephrol* 2008; 19(6):1239-46. doi:10.1681/ASN.2007090993.
- Shioi A, Morioka T, Shoji T, et al. The inhibitory roles of vitamin K in progression of vascular calcification. *Nutrients* 2020; 12(2). doi: 10.3390/nu12020583.
- Halder M, Petsophonsakul P, Akbulut AC, et al. Vitamin K: double bonds beyond coagulation insights into differences between vitamin K1 and K2 in health and disease. *Int J Mol Sci* 2019; 20(4). doi: 10.3390/ijms20040896.
- Wen L, Chen J, Duan L, et al. Vitamin K-dependent proteins involved in bone and cardiovascular health (Review). *Mol Med Rep* 2018; 18(1):3-15. doi: 10.3892/mmr.2018.8940.
- Hauschka PV, Lian JB, Cole DE, et al. Osteocalcin and matrix Gla protein: vitamin K-dependent proteins in bone. *Physiol Rev* 1989; 69(3): 990-1047. doi: 10.1152/physrev.1989.69.3.990.
- Azuma K, Inoue S. Multiple modes of vitamin K actions in aging-related musculoskeletal disorders. *Int J Mol Sci* 2019; 20(11). doi: 10.3390/

- ijms20112844.
16. Viegas CS, Herfs M, Rafael MS, et al. Gla-rich protein is a potential new vitamin K target in cancer: evidences for a direct GRP-mineral interaction. *Biomed Res Int* 2014; 2014:340216. doi: 10.1155/2014/340216.
 17. Pan MH, Maresz K, Lee PS, et al. Inhibition of TNF-alpha, IL-1alpha, and IL-1beta by pretreatment of human monocyte-derived macrophages with menaquinone-7 and cell activation with TLR agonists in vitro. *J Med Food* 2016; 19(7):663-9. doi: 10.1089/jmf.2016.0030.
 18. Shea MK, Booth SL, Massaro JM, et al. Vitamin K and vitamin D status: associations with inflammatory markers in the Framingham Offspring Study. *Am J Epidemiol* 2008; 167(3):313-20. doi: 10.1093/aje/kwm306.
 19. Shea MK, Dallal GE, Dawson-Hughes B, et al. Vitamin K, circulating cytokines, and bone mineral density in older men and women. *Am J Clin Nutr* 2008; 88(2):356-63. doi: 10.1093/ajcn/88.2.356.
 20. Diederichsen ACP, Lindholt JS, Moller S, et al. Vitamin K2 and D in patients with aortic valve calcification: a randomized double-blinded clinical trial. *Circulation* 2022; 145(18): 1387-97. doi: 10.1161/CIRCULATIONAHA.121.057008.
 21. Lees JS, Rankin AJ, Gillis KA, et al. The VIKTORIES trial: a randomized, double-blind, placebo-controlled trial of vitamin K supplementation to improve vascular health in kidney transplant recipients. *Am J Transplant* 2021; 21(10):3356-68. doi: 10.1111/ajt.16566.
 22. Ivanovski O, Szumilak D, Nguyen-Khoa T, et al. Effect of simvastatin in apolipoprotein E-deficient mice with surgically induced chronic renal failure. *J Urol* 2008; 179(4):1631-36. doi: 10.1016/j.juro.2007.11.042.
 23. Afonso P, Auclair M, Boccaro F, et al. LMNA mutations resulting in lipodystrophy and HIV protease inhibitors trigger vascular smooth muscle cell senescence and calcification: role of ZMPSTE24 downregulation. *Atherosclerosis* 2016; 245:200-11. doi: 10.1016/j.atherosclerosis.2015.12.012.
 24. Healy A, Berus JM, Christensen JL, et al. Statins disrupt macrophage *rac1* regulation leading to increased atherosclerotic plaque calcification. *Arterioscler Thromb Vasc Biol* 2020; 40(3): 714-32. doi: 10.1161/ATVBAHA.119.313832.
 25. Son BK, Kozaki K, Iijima K, et al. *Gas6/Axl-PI3K/Akt* pathway plays a central role in the effect of statins on inorganic phosphate-induced calcification of vascular smooth muscle cells. *Eur J Pharmacol* 2007; 556(1-3):1-8. doi: 10.1016/j.ejphar.2006.09.070.
 26. Qiu C, Zheng H, Tao H, et al. Vitamin K2 inhibits rat vascular smooth muscle cell calcification by restoring the *Gas6/Axl/Akt* anti-apoptotic pathway. *Mol Cell Biochem* 2017; 433(1-2): 149-59. doi: 10.1007/s11010-017-3023-z.
 27. Dykun I, Lehmann N, Kalsch H, et al. Statin medication enhances progression of coronary artery calcification: the Heinz Nixdorf Recall Study. *J Am Coll Cardiol* 2016; 68(19):2123-5. doi: 10.1016/j.jacc.2016.08.040.
 28. Vogel LH, Dykun I, Raggi P, et al. High- vs. low-intensity statin therapy and changes in coronary artery calcification density after one year. *J Clin Med* 2023; 12(2). doi: 10.3390/jcm12020476.
 29. Hur DJ, Raymond GV, Kahler SG, et al. A novel MGP mutation in a consanguineous family: review of the clinical and molecular characteristics of Keutel syndrome. *Am J Med Genet A* 2005; 135(1): 36-40. doi: 10.1002/ajmg.a.30680.
 30. O'Young J, Liao Y, Xiao Y, et al. Matrix Gla protein inhibits ectopic calcification by a direct interaction with hydroxyapatite crystals. *J Am Chem Soc* 2011; 133(45):18406-12. doi: 10.1021/ja207628k.
 31. Zebboudj AF, Imura M, Bostrom K. Matrix GLA protein, a regulatory protein for bone morphogenetic protein-2. *J Biol Chem* 2002; 277(6): 4388-94. doi: 10.1074/jbc.M109683200.
 32. Spronk HM, Soute BA, Schurgers LJ, et al. Matrix Gla protein accumulates at the border of regions of calcification and normal tissue in the media of the arterial vessel wall. *Biochem Biophys Res Commun* 2001; 289(2):485-90. doi: 10.1006/bbrc.2001.5996.
 33. Shioi A, Taniwaki H, Jono S, et al. Monckeberg's medial sclerosis and inorganic phosphate in uremia. *Am J Kidney Dis* 2001; 38(4 Suppl 1): S47-49. doi: 10.1053/ajkd.2001.27396.
 34. Ravindran S, Swaminathan K, Ramesh A, et al. Nicorandil attenuates neuronal mitochondrial dysfunction and oxidative stress associated with murine model of vascular calcification. *Acta Neurobiol Exp (Wars)* 2017; 77(1):57-67. doi: 10.21307/ane-2017-036.
 35. Ravindran S, Ramachandran K, Kurian GA. Sodium thiosulfate mediated cardioprotection against myocardial ischemia-reperfusion injury is defunct in rat heart with co-morbidity of vascular calcification. *Biochimie* 2018; 147:80-8. doi: 10.1016/j.biochi.2018.01.004.
 36. Ravindran S, Murali J, Amirthalingam SK, et al. Vascular calcification abrogates the nicorandil mediated cardio-protection in ischemia reperfusion injury of rat heart. *Vascul Pharmacol* 2017; 89:31-8. doi: 10.1016/j.vph.2016.12.004.
 37. Zhu Y, Han XQ, Sun XJ, et al. Lactate accelerates vascular calcification through NR4A1-regulated mitochondrial fission and BNIP3-related mitophagy. *Apoptosis* 2020; 25:321-40. doi: 10.1007/s10495-020-01592-7.
 38. Patel L, Bernard LM, Elder GJ. Sevelamer versus calcium-based binders for treatment of hyperphosphatemia in CKD: a meta-analysis of randomized controlled trials. *Clin J Am Soc Nephrol* 2016; 11(2):232-44. doi: 10.2215/CJN.06800615.
 39. Raggi P, Chertow GM, Torres PU, et al. The ADVANCE study: a randomized study to evaluate the effects of cinacalcet plus low-dose vitamin D on vascular calcification in patients on hemodialysis. *Nephrol Dial Transplant* 2011; 26(4):1327-39. doi: 10.1093/ndt/gfq725.
 40. Grases F, Simonet BM, Vucenik I, et al. Absorption and excretion of orally administered inositol hexaphosphate (IP(6) or phytate) in humans. *Biofactors* 2001; 15(1):53-61. doi: 10.1002/biof.5520150105.
 41. Lopez-Gonzalez AA, Grases F, Monroy N, et al. Protective effect of myo-inositol hexaphosphate (phytate) on bone mass loss in postmenopausal women. *Eur J Nutr* 2013; 52(2):717-26. doi: 10.1007/s00394-012-0377-6.
 42. Joubert P, Ketteler M, Salcedo C, et al. Hypothesis: Phytate is an important unrecognized nutrient and potential intravenous drug for preventing vascular calcification. *Med Hypotheses* 2016; 94: 89-92. doi: 10.1016/j.mehy.2016.07.005.
 43. Ferrer MD, Perez MM, Canaves MM, et al. A novel pharmacodynamic assay to evaluate the effects of crystallization inhibitors on calcium phosphate crystallization in human plasma. *Sci Rep* 2017; 7(1):6858. doi: 10.1038/s41598-017-07203-x.
 44. Perello J, Ferrer MD, Del Mar Perez M, et al. Mechanism of action of SNF472, a novel calcification inhibitor to treat vascular calcification and calciphylaxis. *Br J Pharmacol* 2020; 177(19):4400-15. doi: 10.1111/bph.15163.
 45. Perello J, Joubert PH, Ferrer MD, et al. First-time-in-human randomized clinical trial in healthy volunteers and haemodialysis patients with SNF472, a novel inhibitor of vascular calcification. *Br J Clin Pharmacol* 2018; 84(12):2867-76. doi: 10.1111/bcp.13752.
 46. Raggi P, Bellasi A, Bushinsky D, et al. Slowing progression of cardiovascular calcification with snf472 in patients on hemodialysis: results of a randomized phase 2b study. *Circulation* 2020; 141(9): 728-39. doi: 10.1161/CIRCULATIONAHA.119.044195.
 47. Bushinsky DA, Raggi P, Bover J, et al. Effects of myo-inositol hexaphosphate (SNF472) on bone mineral density in patients receiving hemodialysis: an analysis of the randomized, placebo-controlled CaLIPSO Study. *Clin J Am Soc Nephrol* 2021; 16(5): 736-45. doi: 10.2215/CJN.16931020.
 48. Brandenburg VM, Sinha S, Torregrosa JV, et al. Improvement in wound healing, pain, and quality of life after 12 weeks of SNF472 treatment: a phase 2 open-label study of patients with calciphylaxis. *J Nephrol* 2019; 32(5):811-21. doi: 10.1007/s40620-019-00631-0.
 49. Sinha S, Gould LJ, Nigwekar SU, et al. The CALCIPHIX study: a randomized, double-blind, placebo-controlled, phase 3 clinical trial of SNF472 for the treatment of calciphylaxis. *Clin Kidney J* 2022; 15(1): 136-44. doi: 10.1093/ckj/sfab117.
 50. Chao CT, Yeh HY, Tsai YT, et al. Natural and non-natural antioxidative compounds: potential candidates for treatment of vascular calcification. *Cell Death Discov* 2019; 5:145. doi: 10.1038/s41420-019-0225-z.
 51. Zhou J, Wang H, Shen R, et al. Mitochondrial-targeted antioxidant MitoQ provides neuroprotection and reduces neuronal apoptosis in experimental traumatic brain injury possibly via the Nrf2-ARE pathway. *Am J*

- Transl Res 2018; 10(6):1887-99.
52. Cui L, Zhou Q, Zheng X, et al. Mitoquinone attenuates vascular calcification by suppressing oxidative stress and reducing apoptosis of vascular smooth muscle cells via the Keap1/Nrf2 pathway. *Free Radic Biol Med* 2020; 161:23-31. doi: 10.1016/j.freeradbiomed.2020.09.028.
 53. Rossman MJ, Santos-Parker JR, Steward CAC, et al. Chronic supplementation with a mitochondrial antioxidant (MitoQ) improves vascular function in healthy older adults. *Hypertension* 2018; 71(6):1056-63. doi: 10.1161/HYPERTENSIONAHA.117.10787.
 54. Tharakan B, Holder-Haynes JG, Hunter FA, et al. Alpha lipoic acid attenuates microvascular endothelial cell hyperpermeability by inhibiting the intrinsic apoptotic signaling. *Am J Surg* 2008; 195(2):174-8. doi: 10.1016/j.amjsurg.2007.09.028.
 55. Kim H, Kim HJ, Lee K, et al. Alpha-Lipoic acid attenuates vascular calcification via reversal of mitochondrial function and restoration of Gas6/Axl/Akt survival pathway. *J Cell Mol Med* 2012; 16(2):273-86. doi: 10.1111/j.1582-4934.2011.01294.x.
 56. Himmelfarb J, Ikizler TA, Ellis C, et al. Provision of antioxidant therapy in hemodialysis (PATH): a randomized clinical trial. *J Am Soc Nephrol* 2014; 25(3):623-33. doi: 10.1681/ASN.2013050545.
 57. Safa J, Ardalan MR, Rezaadehsaatlou M, et al. Effects of alpha lipoic acid supplementation on serum levels of IL-8 and TNF-alpha in patient with ESRD undergoing hemodialysis. *Int Urol Nephrol* 2014; 46(8):1633-8. doi: 10.1007/s11255-014-0688-z.
 58. Peng T, Zhuo L, Wang Y, et al. Systematic review of sodium thiosulfate in treating calciphylaxis in chronic kidney disease patients. *Nephrology (Carlton)* 2018; 23(7):669-75. doi: 10.1111/nep.13081.
 59. Zhong H, Liu F, Dai X, et al. Sodium thiosulfate protects human aortic smooth muscle cells from osteoblastic transdifferentiation via high-level phosphate. *Kaohsiung J Med Sci* 2013; 29(11):587-93. doi: 10.1016/j.kjms.2013.04.004.
 60. Chen NX, O'Neill K, Akl NK, et al. Adipocyte induced arterial calcification is prevented with sodium thiosulfate. *Biochem Biophys Res Commun* 2014; 449(1):151-6. doi: 10.1016/j.bbrc.2014.05.005.
 61. Djuric P, Dimkovic N, Schlieper G, et al. Sodium thiosulfate and progression of vascular calcification in end-stage renal disease patients: a double-blind, randomized, placebo-controlled study. *Nephrol Dial Transplant* 2020; 35(1):162-9. doi: 10.1093/ndt/gfz204.
 62. Saengpanit D, Chattranukulchai P, Tumkosit M, et al. Effect of sodium thiosulfate on arterial stiffness in end-stage renal disease patients undergoing chronic hemodialysis (sodium thiosulfate-hemodialysis study): a randomized controlled trial. *Nephron* 2018; 139(3):219-27. doi: 10.1159/000488009.
 63. Granata S, Dalla Gassa A, Tomei P, et al. Mitochondria: a new therapeutic target in chronic kidney disease. *Nutr Metab (Lond)* 2015; 12:49. doi: 10.1186/s12986-015-0044-z.
 64. Cui L, Li Z, Chang X, et al. Quercetin attenuates vascular calcification by inhibiting oxidative stress and mitochondrial fission. *Vascul Pharmacol* 2017; 88:21-9. doi: 10.1016/j.vph.2016.11.006.
 65. Beazley KE, Egtesad S, Nurminkaya MV. Quercetin attenuates warfarin-induced vascular calcification in vitro independently from matrix Gla protein. *J Biol Chem* 2013; 288(4):2632-40. doi: 10.1074/jbc.M112.368639.
 66. Chang XY, Cui L, Wang XZ, et al. Quercetin attenuates vascular calcification through suppressed oxidative stress in adenine-induced chronic renal failure rats. *Biomed Res Int* 2017; 2017:5716204. doi: 10.1155/2017/5716204.
 67. Chen J, Hamm LL, Bundy JD, et al. Combination treatment with sodium nitrite and isoquercetin on endothelial dysfunction among patients with CKD: a randomized phase 2 pilot trial. *Clin J Am Soc Nephrol* 2020; 15(11):1566-75. doi: 10.2215/CJN.02020220.
 68. Suzuki S, Suzuki M, Hanafusa N, et al. Denosumab recovers aortic arch calcification during long-term hemodialysis. *Kidney Int Rep* 2021; 6:605-12. doi: 10.1016/j.ekir.2020.12.002.
 69. Kim H, Lee EJ, Woo S, et al. Effect of denosumab on bone health, vascular calcification, and health-related quality of life in hemodialysis patients with osteoporosis: a prospective observational study. *J Clin Med* 2024;13. doi: 10.3390/jcm13051462.
 70. Al-Aly Z. Arterial calcification: a tumor necrosis factor-alpha mediated vascular Wnt-opathy. *Transl Res* 2008, 151: 233-9. doi: 10.1016/j.trsl.2007.12.005.
 71. Tiong MK, Smith ER, Toussaint ND, et al. Reduction of calciprotein particles in adults receiving infliximab for chronic inflammatory disease. *JBMR Plus* 2021;5:e10497. doi: 10.1002/jbmr4.10497.
 72. Zeng Q, Zhong Y, Yu X. Meta-analysis of the efficacy and safety of sevelamer as hyperphosphatemia therapy for hemodialysis patients. *Ren Fail* 2023;45:2210230. doi: 10.1080/0886022X.2023.2210230.
 73. Lioufas NM, Pascoe EM, Hawley CM, et al. Systematic review and meta-analyses of the effects of phosphate-lowering agents in nondialysis CKD. *J Am Soc Nephrol* 2022;33:59-76. doi: 10.1681/ASN.2021040554.
 74. Hou YC, Zheng CM, Chiu HW, et al. Role of calcimimetics in treating bone and mineral disorders related to chronic kidney disease. *Pharmaceuticals (Basel)* 2022;15. doi: 10.3390/ph15080952.
 75. Mary A, Objois T, Brazier M, et al. Decreased monocyte calcium sensing receptor expression in patients with chronic kidney disease is associated with impaired monocyte ability to reduce vascular calcification. *Kidney Int* 2021;99:1382-91. doi: 10.1016/j.kint.2021.01.026.
 76. Zaslow SJ, Oliveira-Paula GH, Chen W. Magnesium and vascular calcification in chronic kidney disease: current insights. *Int J Mol Sci* 2024;25. doi: 10.3390/ijms25021155.
 77. Ter Braake AD, Smit AE, Bos C, et al. Magnesium prevents vascular calcification in Klotho deficiency. *Kidney Int* 2020, 97: 487-501. doi: 10.1016/j.kint.2019.09.034.
 78. Bressendorff I, Hansen D, Schou M, et al. The effect of magnesium supplementation on vascular calcification in CKD: a randomized clinical trial (MAGICAL-CKD). *J Am Soc Nephrol* 2023;34:886-94. doi: 10.1681/ASN.0000000000000092.
 79. Zhan Y, Zhang R, Li G. Effect of magnesium on vascular calcification in chronic kidney disease patients: a systematic review and meta-analysis. *Ren Fail* 2023;45:2182603. doi: 10.1080/0886022X.2023.2182603.
 80. Zhao L, Yang N, Song Y, et al. Effect of iron overload on endothelial cell calcification and its mechanism. *Ann Transl Med* 2021, 9: 1658. doi: 10.21037/atm-21-5666.
 81. Ye Y, Chen A, Li L, et al. Repression of the antiporter SLC7A11/glutathione/glutathione peroxidase 4 axis drives ferroptosis of vascular smooth muscle cells to facilitate vascular calcification. *Kidney Int* 2022, 102: 1259-75. doi: 10.1016/j.kint.2022.07.034.
 82. Zhu K, Reiser J. ALKBH1 reduces DNA N6-methyladenine to allow for vascular calcification in chronic kidney disease. *J Clin Invest* 2021, 131. doi: 10.1172/JCI150966.
 83. Ouyang L, Su X, Li W, et al. ALKBH1-demethylated DNA N6-methyladenine modification triggers vascular calcification via osteogenic reprogramming in chronic kidney disease. *J Clin Invest* 2021, 131. doi: 10.1172/JCI146985.
 84. Li W, Feng W, Su X, et al. SIRT6 protects vascular smooth muscle cells from osteogenic transdifferentiation via RUNX2 in chronic kidney disease. *J Clin Invest* 2022,132. doi: 10.1172/JCI150051.
 85. Wang S, Tong M, Hu S, et al. The bioactive substance secreted by MSC retards mouse aortic vascular smooth muscle cells calcification. *Biomed Res Int* 2018; 2018:6053567. doi: 10.1155/2018/6053567.
 86. Wang S, Hu S, Wang J, et al. Conditioned medium from bone marrow-derived mesenchymal stem cells inhibits vascular calcification through blockade of the BMP2-Smad1/5/8 signaling pathway. *Stem Cell Res Ther* 2018; 9(1):160. doi: 10.1186/s13287-018-0894-1.
 87. Majesky MW, Dong XR, Regan JN, et al. Vascular smooth muscle progenitor cells: building and repairing blood vessels. *Circ Res* 2011; 108(3):365-77. doi: 10.1161/CIRCRESAHA.110.223800.
 88. Kramann R, Goettsch C, Wongboonsin J, et al. Adventitial MSC-like cells are progenitors of vascular smooth muscle cells and drive vascular calcification in chronic kidney disease. *Cell Stem Cell* 2016; 19(5):628-42. doi: 10.1016/j.stem.2016.08.001.
 89. Angsubhakorn N, Kang N, Fearon C, et al. Contemporary management

- of severely calcified coronary lesions. *J Pers Med* 2022; 12(10). doi: 10.3390/jpm12101638.
90. Bulluck H, McEntegart M. Contemporary tools and devices for coronary calcium modification. *JRSM Cardiovasc Dis* 2022; 11: 20480040221089760. doi: 10.1177/20480040221089760.
91. Secco GG, Ghione M, Mattesini A, et al. Very high-pressure dilatation for undilatable coronary lesions: indications and results with a new dedicated balloon. *EuroIntervention* 2016; 12(3):359-65. doi: 10.4244/EIJY15M06_04.
92. Albiero R, Silber S, Di Mario C, et al. Cutting balloon versus conventional balloon angioplasty for the treatment of in-stent restenosis: results of the Restenosis Cutting Balloon Evaluation Trial (RESCUT). *J Am Coll Cardiol* 2004; 43(6):943-9. doi: 10.1016/j.jacc.2003.09.054.
93. Jujo K, Saito K, Ishida I, et al. Intimal disruption affects drug-eluting cobalt-chromium stent expansion: a randomized trial comparing scoring and conventional balloon predilation. *Int J Cardiol* 2016; 221:23-31. doi: 10.1016/j.ijcard.2016.07.002.
94. Karimi Galougahi K, Patel S, Shlofmitz RA, et al. Calcific plaque modification by acoustic shock waves: intravascular lithotripsy in coronary interventions. *Circ Cardiovasc Interv* 2021; 14(1): e009354. doi: 10.1161/CIRCINTERVENTIONS.120.009354.
95. Brinton TJ, Ali ZA, Hill JM, et al. Feasibility of shockwave coronary intravascular lithotripsy for the treatment of calcified coronary stenoses. *Circulation* 2019; 139(6):834-6. doi: 10.1161/CIRCULATIONAHA.118.036531.
96. Hill JM, Kereiakes DJ, Shlofmitz RA, et al. Intravascular lithotripsy for treatment of severely calcified coronary artery disease. *J Am Coll Cardiol* 2020; 76(22):2635-46. doi: 10.1016/j.jacc.2020.09.603.
97. Saito S, Yamazaki S, Takahashi A, et al. Intravascular lithotripsy for vessel preparation in severely calcified coronary arteries prior to stent placement - primary outcomes from the Japanese Disrupt CAD IV study. *Circ J* 2021; 85(6):826-33. doi: 10.1253/circj.CJ-20-1174.
98. Tepe G, Brodmann M, Werner M, et al. Intravascular lithotripsy for peripheral artery calcification: 30-day outcomes from the randomized disrupt PAD III trial. *JACC Cardiovasc Interv* 2021; 14(12):1352-61. doi: 10.1016/j.jcin.2021.04.010.
99. Lawton JS, Tamis-Holland JE, Bangalore S, et al. 2021 ACC/AHA/SCAI guideline for coronary artery revascularization: a report of the American college of cardiology/American heart association joint committee on clinical practice guidelines. *Circulation* 2022; 145(3):e18-e114. doi: 10.1161/CIR.0000000000001038.
100. Genereux P, Lee AC, Kim CY, et al. Orbital atherectomy for treating de novo severely calcified coronary narrowing (1-year results from the pivotal ORBIT II trial). *Am J Cardiol* 2015; 115(12):1685-90. doi: 10.1016/j.amjcard.2015.03.009.
101. Koster R, Kahler J, Brockhoff C, et al. Laser coronary angioplasty: history, present and future. *Am J Cardiovasc Drugs* 2002; 2(3):197-207. doi: 10.2165/00129784-200202030-00006.
102. Mohandes M, Rojas S, Moreno C, et al. Excimer laser in percutaneous coronary intervention of device uncrossable chronic total and functional occlusions. *Cardiovasc Revasc Med* 2020; 21(5):657-60. doi: 10.1016/j.carrev.2019.08.022.
103. Lee T, Shlofmitz RA, Song L, et al. The effectiveness of excimer laser angioplasty to treat coronary in-stent restenosis with peri-stent calcium as assessed by optical coherence tomography. *EuroIntervention* 2019; 15(3):e279-e88. doi: 10.4244/EIJ-D-18-00139.
104. Kawashima H, Serruys PW, Hara H, et al. 10-year all-cause mortality following percutaneous or surgical revascularization in patients with heavy calcification. *JACC Cardiovasc Interv* 2022; 15(2):193-204. doi: 10.1016/j.jcin.2021.10.026.
105. Naggara O, Touze E, Beyssen B, et al. Anatomical and technical factors associated with stroke or death during carotid angioplasty and stenting: results from the endarterectomy versus angioplasty in patients with symptomatic severe carotid stenosis (EVA-3S) trial and systematic review. *Stroke* 2011; 42(2):380-8. doi: 10.1161/STROKEAHA.110.588772.

(Edited by Liang-Jun Gu)